



Antimicrobial activity of phytoextracts on opportunistic oral bacteria, yeast and bacteria from probiotics

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Developed experimental assays enable us to compare the antimicrobial activity of herbal medicinal drugs on *Lactobacillus* and *Bacillus* strains probiotics, which have been claimed to possess the ability of suppressing the growth of various oral pathogens. In the treatment of periodontal disease it is advisable to use a comprehensive approach which would include the application of herbal remedies and probiotics. The combination of such effects may be a new approach in dentistry due to their complementary antimicrobial activity. In this study, we researched antimicrobial effects of herbal medicinal drugs (tinctures of some medical plants, solutions Rotocanum and Chlorophyllipt) against collection strains and clinical strains isolated from the oral cavity of patients with periodontitis, and probiotic strains *Bacillus subtilis* UKM B-5007 and *Bacillus licheniformis* UKM B-5514 that are part of the active base of probiotic Biosporin (Ukraine), *Bacillus clausii* from the probiotic Normaflore (Hungary), as well as the strains *Lactobacillus* spp. – from probiotic *Lactobacterinum* (Biopharma, Ukraine). For investigation, the standard agar dilution method was used in modification with glass cylinders. The results of the research showed that among the studied herbal medicinal remedies, tinctures of *Eucalyptus viminalis*, *Mentha piperita* and Chlorophyllipt had the strongest antimicrobial activity. Probiotic strains are also sensitive to herbal tinctures (except the tincture of wormwood), which indicate the possibility of only consecutive usage (with an interval of time) of herbal remedies and probiotics in combination therapy in the treatment of periodontal diseases.

Keywords: antimicrobial activity; herbal medicinal remedies; probiotics; periodontal diseases.

Introduction

Oral health influences the general quality of life, and poor oral health is linked to chronic conditions and systemic diseases. Dental caries, gingivitis and periodontitis are prominent oral disorders. The current understanding is that the etiology of oral diseases is multifactorial but, in many cases, it includes a pathogenic response to bacterial and *Candida* infection (Radulovic et al., 2013; Vorobets & Rivis, 2017). Periodontitis is a common and widespread disease, which occurs due to pathogenic microbial infection established within the gingival sulcus. Treatment of patients with periodontal diseases should include both local and general therapy. It must be based on the understanding of the mechanisms of action of the pharmaceutical or prophylactic agents, and be used to be effective and at the same time safe for the patient. Currently, the drugs of plant origin correspond to such criteria, because active compounds in their composition can act bacteriostatically and bactericidally (Rios et al., 2005; Kačaniová et al., 2014; Hleba et al., 2016). Another group of medicines – probiotics have relatively recently started to be used for periodontal treatment. Probiotics are commonly bacteria from genera *Lactobacillus* (including strains *L. salivarius* and *L. plantarum*) and *Bifidobacterium* (including strains *B. bifidum*, *B. longum*) or others, which

could be beneficial not only to the digestive system, but to oral health too (Mishra et al., 2014; Alok et al., 2017).

The mechanisms of probiotic action are mainly unknown but the inter-microbial species interactions are supposed to play a key role in this together with their immunostimulatory effects. The proposed mechanisms of action of probiotics on oral health correspond to those shown in studies of the gastrointestinal tract. These are aid in synthesis of vitamins B, and K, and also in the breakdown of bile salts, aid in enhancing innate and acquired immunity, and assistance in inhibition of pro-inflammatory mediators (Alok et al., 2017). It is known that oral diseases, including periodontitis, are often accompanied by qualitative and quantitative changes in the microbiota (Rivis et al., 2012; Curtis, 2014). This makes probiotics an alternative means of correction of microbiota of the mouth and of reducing the number of opportunistic pathogens. On the other hand, there is evidence that the occurrence of candidiasis of the oral cavity and its chronic form is caused not only by changes in the balance of microorganisms in the mouth, but also by the influence of intestinal microbiota (Zlatkina et al., 2001).

Therefore, a complex treatment should be performed targeting the mouth and lower parts of the gastrointestinal tract. Dysbacteriosis is not only excessive growth of pathogenic microbes in the gut, but also

involves the overall decline of the immune defence of the body (Alok et al., 2017) At the same time, it can involve such diseases as atopic dermatitis, eczema, thirst, bronchial asthma, food allergy in children. To restore normal intestinal microbiota, probiotics can be used, which are often composed of bifidobacteria and lactobacilli, which are able to show antagonism against pathogenic and opportunistic microorganisms. The results of our previous studies indicate that some industrial strains of spore bacteria that are used as a basis for probiotic products show high antagonistic activity against strains of microorganisms isolated from “periodontal pockets” (Rivis et al., 2013). However, from the clinical practitioner’s point of view, direct recommendations for the use of probiotics cannot yet be given (Curtis, 2014). Beside this, there is a lack of information regarding the contributions of probiotics in oral health and their compatible use with plant origin remedies (Safronova, 2009; Shipradeep, 2012). Our research is focused on the determining of the antimicrobial properties of Tinctura Eucalypti, Tinctura Calendulae, Tinctura Menthae piperitae, Tinctura Absinthii, and also Tinctures Chlorophyllipt and Rotocanum against collection and clinical strains isolated from the oral cavity of patients with periodontitis and against probiotic strains that are part of the active base of probiotics Biosporin, Lactobacterin and Normaflora.

Material and methods

The effect of herbal medicinal products such as Rotocanum, Chlorophyllipt, Tinctura Calendulae, Tinctura Eucalypti, Tinctura Menthae piperitae, Tinctura Absinthii was investigated on collection and clinical isolated strains of microorganisms. The general chemical profiles of the extracts and their pharmacological effects against oral cavity diseases are summarized in Table 1.

To determine the antimicrobial activity of the herbal medicinal remedies, as test cultures we used bacteria from the American Type Culture Collection, USA: *Escherichia coli* ATCC 25922, *Staphylococcus*

aureus ATCC 25923, *Enterococcus faecalis* ATCC 29212, and yeast *Candida albicans* ATCC 885-653; clinical strains of bacteria: *Staphylococcus aureus*, *Streptococcus salivarius*, *Enterobacter* sp., *Neisseria* sp.; yeast *Candida albicans*, isolated from the oral cavity of periodontitis patients; and bacteria *Bacillus subtilis* UKM B-5007 and *Bacillus licheniformis* UKM B-5514, isolated from probiotic Biosporin (Biopharma, Ukraine), and *Bacillus clausii* from the probiotic Normaflora (Manufacturer: Uniter Laboratory, France; holder of Sanofi-aventis S.p.A., Hungary trade license), as well as the strains *Lactobacillus* spp. – from probiotic Lactobacterinum (Biopharma, Ukraine). Positive control were prepared with the same solvents, which were used to prepare the plant Tinctures. The antiseptic drug Decasan (Solution of decamethoxine dihydrochloride 0.02% by weight in water with sodium chloride, Yuria-Pharm Ltd.) also has antimicrobial properties.

The agar diffusion method as adapted earlier using glass cylinders (Vorobets & Yavorska, 2016) was used. From the daily culture of microorganisms, a suspension was made in a sterile physiological solution, and every suspension was adjusted to equal 0.5 McFarland standard. Each cup of Muller-Hinton agar was filled with 0.1 ml of a bacterial suspension. The cups were dried at room temperature for an hour. Then on the culture medium with tweezers we carefully arranged sterile glass cylinders, into which 0.1 ml of the substance was contributed. Antimicrobial activity was judged by the presence and size of the growth zone of the studied microorganisms around the cylinder with the extract. To determine the antimicrobial activity of the examined samples, the following scale was used: diameter of the growth retardation zone more than 20 mm highly sensitive, 10–20 mm – sensitive, up to 10 mm – moderately sensitive. Ethanol of various concentrations was used as solvent control. All tests were performed at least three times.

Results of laboratory tests were processed by methods of variation statistics with the calculation of averages (M) and their error (m), the criterion of authenticity difference is determined using Fisher tables and adapted to the Medical Research simplified tables.

Table 1
Characteristics of Drugs Used

Latin name of the drug. Manufacturer	Active substance. Extractant	Pharmaceutical composition, basic active substances	Mode of action
Tinctura Eucalypti. LLC “DKP “Pharmaceutical Factory”, Zhytomyr, Ukraine	<i>Eucalypti viminalis</i> folia Extractant 70% ethanol	Eucalyptus leaves contain essential oil (3%), flavonoids, tannins, ellagic acid, resins and waxes	Used to treat stomatitis, gangrenous pulpitis, inflammation of the oral mucosa; in otorhinolaryngology for the treatment of sore throat, chronic rhinitis and pharyngitis. In surgery, used to treat of abscesses, osteomyelitis, purulent mastitis, open fractures, burns and frostbite (Shulga, 2011)
Tinctura Calendulae. LLC “DKP “Pharmaceutical Factory”, Zhytomyr, Ukraine	<i>Calendulae officinalis</i> floridis Extractant 70% ethanol	Saponins, tannins, flavonoid and carotenoid glycosides, organic acids, essential oil, carbohydrates resins, saponins, organic acids	In tincture form, taken internally, promotes healing of ailments in the digestive tract; also has anti-inflammatory and antibacterial activity (Shulga, 2011). Calendula's high molecular weight polysaccharides stimulate the immune system activity; have anti-inflammatory, antimicrobial, and moderate choleric effect
Tinctura Menthae piperitae. PrAT Pharmaceutical Factory “Viola”, Zaporozhye, Ukraine	<i>Menthae piperitae</i> folia Extractant 90% ethanol	Glycoside tropolin, essential oils, menthol, potassium sulfate, phytosterols, ascorbic acid, sugar, starch, mucilage, pectin, pigment sorbuzin and others	Normalizes heart rate, improves circulation, relieves spasms of blood vessels and expands them, lowers blood pressure, and relieves stress. In dentistry is widely used as a component of menthol mouthwash, toothpastes and powders, solutions for rinsing the oral cavity in purpose of breath freshening
Tinctura Absinthii or Absinthii tinctura. PrAT Pharmaceutical Factory “Viola”, Zaporozhye, Ukraine	<i>Artemisiae absinthii</i> herba Extractant 70% ethanol	Bitternesses	Has anti-inflammatory, antiseptic, anti-ulcer effect; used for the treatment of gingivitis, periodontitis; stimulates appetite, activates all stages of digestion
Chlorophyllipt. Corporation “Arterium”, JSC “Halychpharm”, Lviv, Ukraine	Extractum chlorophyllipti spissum Extractant 96% ethanol	Chlorophylls mixture of eucalyptus leaves	Has antibacterial (bacteriostatic and bactericidal) activity against staphylococci and antiseptic and anti-inflammatory activity; used to rinse the mouth at the stage of initial periodontal treatment; for local utilization (Shulga, 2011)
Rotocanum. State Enterprise “Experimental Plant of Medicines IBONH NAS of Ukraine”, Kyiv, Ukraine	A mixture of liquid extracts (2 : 1 : 1): <i>Matricariae recutitae</i> extractum fluidum + <i>Calendulae</i> extractum fluidum + <i>Millefolii</i> extractum fluidum. Extractant 40% ethanol	–	Injected into the dental pockets in patients with parodontosis; in cases of stomatitis used as oral applications or baths (Shulga, 2011)

Results

The antimicrobial activity of the herbal medicinal remedies was tested *in vitro* (Table 2). Tinctura Calendulae showed antimicrobial effect on

E. coli, *S. aureus*, *E. faecalis*, and probiotic strains. Moreover, such activity was greater than that of the antiseptic Decasan. However, it showed no or low antifungal activity and effects on bacteria of genus *Neisseria* and *Enterobacter* compared to controls. Research has shown

that Tinctura Eucalypti has high antimicrobial activity against Gram-positive, Gram-negative microorganisms, and probiotic strains. The diameters of zones of growth retardation were the highest for staphylococci (up to 30 mm), streptococci (up to 30 mm) and *B. clausii* (up to 40 mm) and significantly exceeded the effect of Decasan on test culture. The tincture did not reveal a pronounced antifungal effect compared to the control. Tinctura Menthae piperitae produces using 90% ethanol and Chloro-

phyllipt 96% one, but despite the presence of the extractant revealed high antimicrobial activity of these drugs against Gram-positive microorganisms, so that diameters of growth were in the range of 20 to 40 mm. These indicators significantly exceeded the effect of Decasan on test cultures. The Tinctura of peppermint inhibited the growth of Gram-negative microorganisms too. Phytopreparations of mint and Chlorophyllipt also have a high anti-candidal effect.

Table 2
Antimicrobial activity of plant medicinal drugs to test-bacteria

Plant medicinal remedies	<i>C. albicans</i>	<i>S. aureus</i>	<i>E. faecalis</i>	<i>E. coli</i>	<i>C. albicans</i>	<i>S. aureus</i>	<i>Enterobacter</i> sp.	<i>Neisseria</i> sp.	<i>S. salivarius</i>	<i>B. subtilis</i>	<i>B. clausii</i>	<i>Lactobacillus</i> sp. (Lactobacterinum)
	ATCC 885-653	ATCC 25923	ATCC 29212	ATCC 25922						<i>B. subtilis</i> UKM B-5007, <i>B. licheniformis</i> UKM B-5514 (Biosporin)		
Tinctura Absinthii	18.0±1.6	8.8±2.0	16.5±0.6*	10.3±0.3	17.6±0.4	12.8±2.9	13.3±1.0	6.7±1.9	19.6±0.9**	0	21.3±1.3*	18.3±1.0
Tinctura Calendulae	19.5±1.8	16.5±0.5**	24.5±0.6*	14.7±2.7	15.9±3.3	13.1±1.9	12.5±0.5*	6.0±0.4	18±2.1*	23.5±0.6***	28.6±1.3***	25.0±1.7**
Tinctura Eucalypti 70% ethanol	18.0±0.0	29.3±2.4***	30.0±0.0*	18.0±1.7	20.2±0.4	30.0±1.5***	17.3±0.7*	21.3±0.3***	28.4±1.1***	26.0±1.9***	37.6±1.0***	28.7±0.7***
Rotocanum 40% ethanol	18.3±1.1	10.0±1.9	0.0±0.0	12.5±1.6	18.9±0.7	9.4±1.4	13.3±0.3	10.5±1.6	10.0±2.5	0.0±0.0	11.3±3.6	17.1±1.4
90% ethanol	10.3±2.4	12.8±0.6**	15.5±0.6*	10.0±0.0*	11.8±0.8*	13.3±1.0***	10.0±0.0***	14.4±1.9*	18.2±1.0***	13.7±0.3***	16.3±1.6**	15.0±2.7**
Tinctura Menthae piperitae	8.1±1.9	5.1±1.9	0.0±0.0	0.0±0.0	6.1±1.7	0.0±0.0	0.0±0.0	6.0±0.3	0.0±0.0	0.0±0.0	0.0±0.0	0.0±0.0
90% ethanol	35.7±2.7**	25.8±1.3**	29.0±1.3*	39.3±0.7***	39.5±0.5***	25.7±1.3**	27.0±0.7*	21.0±1.0**	36.3±0.7*	26.4±4.4***	40.5±1.7***	30.3±0.3*
90% ethanol	24.8±1.1	20.5±1.1	14.0±2.7	21.5±1.2	21.9±0.9	19.3±1.1	19.3±2.4	16.5±0.7	26.3±2.1	0.0±0.0	25.0±1.6	20.2±4.5
Chlorophyllipt	30.8±2.1	40.0±1.6***	37.5±1.7*	21.7±1.0	23.3±1.0	34.8±2.5***	19.3±2.1	32.0±1.2***	35.0±2.7	37.2±1.7***	40.4±1.2***	39.3±0.7**
96% ethanol	26.9±0.9	22.4±0.8	12.0±2.5	23.3±2.4	24.5±1.0	19.2±0.7	23.0±1.7	18.4±0.6	30.5±0.5	6.0±0.3	25.8±0.8	26.0±0.0

Note: mathematical reliability * – P < 0.05, ** – P < 0.01, *** – P < 0.001; the reliability of the difference between the plant medicinal remedies and the corresponding solution of alcohol P: Tinctura Absinthii, Tinctura Calendulae, Tinctura Eucalypti – 70% ethanol; Rotocanum – 40% ethanol; Tinctura Menthae piperitae – 90% ethanol; Chlorophyllipt – 96% ethanol.

Sensitive to the Tinctura Absinthii were *S. salivarius* (19.6±0.9 mm), *E. faecalis* (16.5 ± 0.6 mm), and *B. clausii* (21.3 ± 1.3 mm) from the probiotic Normaflore, as well as *C. albicans*, *S. aureus* and *Lactobacillus* sp., but ethanol also had an effect on these microorganisms. Tinctura Absinthii did not affect the growth of Gram-negative microorganisms, and species of probiotic Biosporin. The drug Rotocanum, which contains in its composition extracts of chamomile, calendula and yarrow (*Calendula officinalis* floridis + *Chamomilla recutitae* floridis + *Achillea millefolii* herbae) and is used as applications or mouthparts baths for patients with periodontitis and stomatitis, did not significantly affect the microorganisms which have been studied. The largest growth retardation regions were observed for streptococci (15.5 ± 0.6 mm for *E. faecalis* ATCC 29212 and 18.2 ± 1.0 mm *S. salivarius*), staphylococci (12–14 mm), and *Neisseria* sp. (14.4 ± 1.9 mm).

It should also be noted that all remedies as extractants containing ethanol are known as disinfectants that also have antimicrobial properties. It was found that Tinctura Absinthii had no effect on bacteria that are the active Biosporin basis. However, it affected the most opportunistic microbes taken in the experiment.

Discussion

Detailed analysis of the impact of drugs on different types of microorganisms made it possible to find that no drug that would act the same way or have no effect on the growth of microorganisms. Thus, the results obtained show that use of herbal drugs in dentistry is promising, in particular to eliminate infectious processes in the mouth and suppression of vital activity of pathogenic and opportunistic microorganisms.

The subject of antimicrobial properties of essential oils and extracts from *Artemisia* L. species is very widely discussed (Erel et al., 2007; Massiha et al., 2013). Cha (2007) investigated the chemical composition and antibacterial activity of *Artemisia iwayomogi* essential oil against oral bacteria. The essential oil of the plant exhibited strong inhibitory effect against all obligate anaerobic bacteria tested, while its major compounds demonstrated various degrees of growth inhibition (Cha, 2007). In our study, ethanolic extract of *Artemisia absinthii* herba showed moderate effect on the investigated strains. Sensitive to the Tinctura Absinthii were mainly Gram-positive oral bacteria and yeast while it did not affect the growth of Gram-negative microorganisms and probiotic

strains *B. subtilis* UKM B-5007, *B. licheniformis* UKM B-5514. Moslemi et al. (2012) showed that topical application of *A. absinthium* extract on the infected wound sites in rat models produced significant antibacterial activity against *S. aureus*.

In the present study, Tinctura Calendulae showed antimicrobial effect on most Gram-positive, Gram-negative microorganisms and probiotic strains. Szakiel et al. (2008) found that oleanic acid isolated from marigold (*Calendula officinalis*) inhibited bacterial growth and survival, influenced cell morphology and enhanced the autolysis of Gram-positive bacteria, suggesting that bacterial envelopes are the target of its activity. Essential oil from *Calendula officinalis* was effective against 23 clinical fungi strains tested (Gazim et al., 2008). Other experimental data obtained showed that methanol extract of *C. officinalis* petals exhibited better antibacterial activity than ethanol extract. However, both extracts showed high antifungal activity in comparison with fluconazole (Efstratiou et al., 2012). In our research Tinctura Calendulae has shown no or low antifungal activity in comparison with control.

Among the studied herbal medicinal products, the strongest antimicrobial activity was found in Tinctura Eucalypti, Tinctura Menthae piperitae and Chlorophyllipt. Eucalyptus oil, which is known for its antibacterial, antiviral (Cermelli et al., 2007; Astani et al., 2009) and antifungal (Ashour, 2008) properties, and has a long history of use for the treatment of colds, flu, rhinitis, sinusitis and other respiratory tract diseases. The obtained results have shown that essential oils of the leaves of *E. globulus* have antimicrobial activity against Gram-negative bacteria (*E. coli*) as well as Gram-positive bacteria (*S. aureus*) (Bachir & Benali, 2012). The effectiveness of eucalyptus oils on caries and periodontitis pathogens was also investigated. In particular, essential oils of *Eucalyptus camaldulensis* showed antibacterial activity against *Streptococcus mutans* and significantly retard its biofilm formation (Rasooli et al., 2009). Takarada et al. (2004) earlier showed that eucalyptus oil inhibited the growth of the following oral bacteria: *Porphyromonas gingivalis*, *Actinobacillus actinomycetemcomitans*, *Fusobacterium nucleatum*, *Streptococcus mutans*, *Streptococcus sorbinus* and also inhibited the adhesion of *S. mutans* (Takarada et al., 2004). The antifungal effect of eucalyptus essential oils has been investigated. Ashour et al. (2008) showed that essential oils of *E. sideroxylon* and *E. torquata* generally exhibited moderate to high antifungal activities against *Candida albicans*, *A. flavus* and *A. niger* (Ashour et al., 2008). Agarwal et al. (2008) inves-

tingated the ability of eucalyptus essential oil to suppress the formation of *C. albicans* biofilm. Takahashi et al. (2004) reported that extracts of *Eucalyptus globulus*, *E. maculata* and *E. viminalis* significantly inhibited the growth of six Gram-positive bacteria (*Staphylococcus aureus*, MRSA, *Bacillus cereus*, *Enterococcus faecalis*, *Alicyclobacillus acidoterrestris*, *Propionibacterium acnes*), and of a fungus (*Trichophyton mentagrophytes*), but they did not show strong antibacterial activity against Gram-negative bacteria (*Escherichia coli*, *Pseudomonas putida*) (Takahashi et al., 2004). In our findings, Gram-negative bacteria showed lower sensitivity to Tinctura Eucalypti than Gram-positive bacteria.

Our results showed that the Tinctura of peppermint inhibited the growth of Gram-positive, Gram-negative microorganisms and had a high anti-candidal effect. Caretto et al. (2010) also showed antimicrobial activity of hydroalcoholic extract of *Mentha piperita* L. against *Candida* spp. (*C. albicans*, *C. tropicalis* and *C. glabrata*). Shalayel et al. (2017) showed the potential antibacterial activity for *Mentha piperita* extracts against MDR *S. pyogenes*, *E. faecalis*, MRSA, MRSE and carbapenem-resistant *E. coli*, and *Klebsiella pneumonia* clinical isolates. Sujana et al. (2013) earlier demonstrated that the organic extracts of the leaves of the plant (*Mentha piperita* L.) possessed strong antibacterial activity against a range of pathogenic bacteria, such as *Bacillus subtilis*, *Streptococcus pneumonia*, *Staphylococcus aureus*, *Escherichia coli*, *Proteus vulgaris* and *Klebsiella pneumonia* (Sujana et al., 2013). The mint leaf methanolic extract showed considerable antimicrobial activity against human oral pathogens, such as: *Escherichia coli*, *Acinetobacter* sp., *Staphylococcus aureus* and two fungi such as *Candida albicans*, *C. glabrata* (Pramila et al., 2012). Miloš Nikolić et al. reported antimicrobial activity of essential oil of peppermint *Mentha piperita* against pathogenic microorganisms isolated from the oral cavity (8 bacteria and 58 *Candida* sp.) and referent strains (Nikolić et al., 2013).

According to usage instruction, Chlorophyllipt inhibits staphylococci infections that are resistant to antibiotics and is used in washing, rinsing, lotion, wet tampons and douching, and in dental stomatitis to treat gangrenous pulpitis, abscesses, boils, inflammation of the oral mucosa. Our results suggested its antibiotic activity not only against collection and clinical strains of Gram-positive and Gram-negative bacteria, but also against investigated probiotic strains. By investigation of the antagonistic activity of probiotics it was established that bacteria, being the background of the biopreparations, have demonstrated different levels of suppression effect on various strains of test-cultures.

Conclusions

It was confirmed that each of the proposed health benefits should be studied separately for each probiotic bacterial strain, especially if the treatment protocol stipulates the use of drugs with herbal remedies. At the same time, all probiotic strains proved to be sensitive to the action of plant extracts, indicating the possibility of consecutive use (with an interval of time) of herbal remedies and probiotics in combination therapy for the treatment of periodontal disease. Screening herbal medicinal products according to their activity on the *Lactobacillus* and *Bacillus* strains of probiotics could precede the clinical efficacy studies for adjunct treatment with both in treatment of periodontal infections.

While BAS of herbal medicines suppress the vital activity of pathogenic and opportunistic microorganisms or destroy them, probiotics create conditions for the resumption of normal microbiota. Alternating drugs (of plant origin and probiotics) in the treatment of periodontal disease will prevent the formation of resistant strains of oral microbiota, and will not disturb the biological balance in the oral cavity, and, therefore, will provide recovery of the patient's health.

References

Agarwal, V., Lal, P., & Pruthi, V. (2008). Prevention of *Candida albicans* biofilm by plant oils. *Mycopathologia*, 165(1), 13–19.

Alok, A., Singh, I. D., Singh, S., Kishore, M., Jha, P. C., & Iqbal, M. A. (2017). Probiotics: A new era of biotherapy. *Advanced Biomedical Research*, 6(1), 31.

Ashour, H. M. (2008). Antibacterial, antifungal and anticancer activities of volatile oils and extracts from stems, leaves, and flowers of *Eucalyptus sideroxylon* and *Eucalyptus torquata*. *Cancer Biology and Therapy*, 7(3), 399–403.

Astani, A., Reichling, J., & Schnitzler, P. (2009). Comparative study on the antiviral activity of selected monoterpenes derived from essential oils. *Phytotherapy Research*, 24(5), 673–679.

Bachir, R. G., & Benali, M. (2012). Antibacterial activity of the essential oils from the leaves of *Eucalyptus globulus* against *Escherichia coli* and *Staphylococcus aureus*. *Asian Pacific Journal of Tropical Biomedicine*, 2(9), 739–742.

Caretto, C. F. P., Junqueira, J. C., Almeida, R. B. A., Furlan, M. R., & Jorge, A. O. C. (2010). Antimicrobial activity of *Mentha piperita* L. against *Candida* spp. *Brazilian Dental Science*, 13(1/2), 4–9.

Cermelli, C., Fabio, A., Fabio, G., & Quaglio, P. (2007). Effect of eucalyptus essential oil on respiratory bacteria and viruses. *Current Microbiology*, 56(1), 89–92.

Cha, J.-D. (2007). Chemical composition and antibacterial activity against oral bacteria by the essential oil of *Artemisia ivayomogi*. *Journal of Bacteriology and Virology*, 37(3), 129–136.

Curtis, M. A. (2014). Periodontal microbiology – The lid's off the box again. *Journal of Dental Research*, 93(9), 840–842.

Efstratiou, E., Hussain, A. I., Nigam, P. S., Moore, J. E., Ayub, M. A., & Rao, J. R. (2012). Antimicrobial activity of *Calendula officinalis* petal extracts against fungi, as well as Gram-negative and Gram-positive clinical pathogens. *Complementary Therapies in Clinical Practice*, 18(3), 173–176.

Erel, Ş. B., Yavaoğlu, N. Ü. K., & Zeybek, U. (2007). Antimicrobial activities of six artemisia species of West Anatolia. *Planta Medica*, 73(9), 131.

Gazim, Z. C., Rezende, C. M., Fraga, S. R., Estivalet Svidzinski, T. I., & Garsia Cortez, D. A. (2008). Antifungal activity of the essential oil from *Calendula officinalis* L. (Asteraceae) growing in Brazil. *Brazilian Journal of Microbiology*, 39(1), 61–63.

Hleba, L., Kompas, M., Hutková, J., Rajtar, M., Petrová, J., Čuboň, J., Kántor, A., & Kačániová, M. (2016). Antimicrobial activity of crude ethanolic extracts from some medicinal mushrooms. *Journal of Microbiology, Biotechnology and Food Sciences*, 5(1), 60–63.

Kačániová, M., Vuković, N., Horska, E., Salamov, I., Bobkova, A., Hleba, L., Mellen, M., Vatlak, A., Petrova, J., & Bobko, M. (2014). Antibacterial activity against *Clostridium* genus and antiradical activity of the essential oils from different origin. *Journal of Environmental Science and Health, Part B*, 49(7), 505–512.

Massiha, A., Majid Khoshkholgh-Pahlavian, M., Issazadeh, K., Bidarigh, S., & Zarabi, S. (2013). Antibacterial activity of essential oils and plant extracts of *Artemisia (Artemisia annua L.) in vitro*. *Zahedan Journal of Research in Medical Sciences*, 15(6), 14–18.

Mishra, R., Tandon, S., Rathore, M., & Banerjee, M. (2014). Antimicrobial and plaque inhibitory potential of herbal and probiotic oral rinses in children: A randomized clinical trial. *Indian Journal of Dental Research*, 25(4), 485–492.

Moslemi, H. R., Hoseinzadeh, H., Badouei, M. A., Kafshdoudzan, K., & Fard, R. M. (2012). Antimicrobial activity of *Artemisia absinthium* against surgical wounds infected by *Staphylococcus aureus* in a rat model. *Indian Journal of Microbiology*, 52(4), 601–604.

Nikolić, M., Glamočlija, J., Ćirić, A., Marković, T., Marković, D., Perić, T., & Soković, M. (2013). Hemijski sastav i antimikrobna aktivnost etarskog ulja pitome nane (*Mentha piperita* L.). *Lekovite Sirovane*, 33, 63–72.

Pramila, D. M., Xavier, R., Marimuthu, K., Kathiresan, S., Khoo, M. L., Senthilkumar, M., Sathya, K., & Sreeramanan, S. (2012). Phytochemical analysis and antimicrobial potential of methanolic leaf extract of peppermint (*Mentha piperita*: Lamiaceae). *Journal of Medicinal Plants Research*, 6(2), 331–335.

Radulovic, N. S., Blagojevic, P. D., Stojanovic-Radic, Z. Z., & Stojanovic, N. M. (2013). Antimicrobial plant metabolites: Structural diversity and mechanism of action. *Current Medicinal Chemistry*, 20(7), 932–952.

Rios, J. L., & Recio, M. C. (2005). Medicinal plants and antimicrobial activity. *Journal of Ethnopharmacology*, 100, 80–84.

Rasooli, I., Shayegh, S., & Astaneh, S. D. A. (2009). The effect of *Mentha spicata* and *Eucalyptus camaldulensis* essential oils on dental biofilm. *International Journal of Dental Hygiene*, 7(3), 196–203.

Rivis, O. Y., Krivtsova, M. V., Nikolaichuk, V. I., Semenova, G. M., & Barani, E. A. (2012). Mikroflora rotovo' porozhnyh ljudej z zapal'nymy zahvorjuvannjamy tkanyny parodontu v Uzhgorod'skomu rajoni [Microflora of the oral cavity of people with inflammatory diseases of periodontal tissue in Uzhgorod area]. *Bulletin of Problems in Biology and Medicine*, 1(3), 121–124 (in Ukrainian).

Rivis, O. Y., Krivtsova, M. V., & Nikolaichuk, V. I. (2013). Antagonistic activity of *Bacillus* probiotics against bacteria isolates of oral cavity of patients with periodontitis. *Visnyk of Dnipropetrovsk University. Biology, Medicine*, 4(1), 10–13.

Safironova, L. A., Osadchaya, A. I., Avdeyeva, L. V., & Ilyash, V. M. (2009). Vliyanie fitokompozicij na biologicheskuju aktivnost' probioticheskih shtammov *Bacillus subtilis*. [Influence of phytocompositions on biological activity of *Bacillus subtilis* probiotic strains]. *Likarska Sprava*, 3–4, 68–74 (in Russian).

Shalayel, M. H. F., Asaad, A. M., Qureshi, M. A., & Elhussein, A. B. (2017). Anti-bacterial activity of peppermint (*Mentha piperita*) extracts against some emerging multi-drug resistant human bacterial pathogens. *Journal of Herbal Medicine*, 7, 27–30.

- Shipra Deep, K. S., Khare, R. S., Ojha, S., Kundu, K., & Kundu, S. (2012). Development of probiotic candidate in combination with essential oils from medicinal plant and their effect on enteric pathogens: a review. *Gastroenterology Research and Practice*, 2012, 1–6.
- Shulga, L. I. (2011). Fitopreparaty v stomatologii: Suchasnyj stan ta perspektyvy stvorennja [Herbal remedies in dentistry: Current status and perspectives for production]. *Clinical Pharmacy, Pharmacotherapy and Medical Standardization*, 3–4, 151–156 (in Ukrainian).
- Sujana, P., Sridhar, T. M., Josthna, P., & Naidu, C. V. (2013). Antibacterial activity and phytochemical analysis of *Mentha piperita* L. (Peppermint) – An important multipurpose medicinal plant. *American Journal of Plant Sciences*, 4(1), 77–83.
- Szakiel, A., Ruszkowski, D., Grudniak, A., Kurek, A., Wolska, K. I., Doligalska, M., & Janiszowska, W. (2008). Antibacterial and antiparasitic activity of oleanolic acid and its glycosides isolated from marigold (*Calendula officinalis*). *Planta Medica*, 74(14), 1709–1715.
- Takahashi, T., Kokubo, R., & Sakaino, M. (2004). Antimicrobial activities of eucalyptus leaf extracts and flavonoids from *Eucalyptus maculata*. *Letters in Applied Microbiology*, 39(1), 60–64.
- Takarada, K., Kimizuka, R., Takahashi, N., Honma, K., Okuda, K., & Kato, T. (2004). A comparison of the antibacterial efficacies of essential oils against oral pathogens. *Oral Microbiology and Immunology*, 19(1), 61–64.
- Vorobets, N., & Ravis, O. (2017). Aktual'nist' ta perspektyvy vykorystannja likars'kyh roslyn dlja likuvannja kandydozu rotovoi' porozhnyny [Relevance and perspectives of using medicinal plants for the treatment of oral candidiasis]. *Bulletin of Problems in Biology and Medicine*, 135, 22–32 (in Ukrainian).
- Vorobets, N. M., & Yavorska, H. V. (2016). Modifications of agar diffusion method to determination of the antimicrobial effect of the herbal medicinal products. *Ukrainian Biopharmaceutical Journal*, 43, 80–84.
- Zlatkina, A. R., Isakov, V. A., & Ivanikov, I. O. (2001). Kandidoz kishchechnika kak novaja problema gastrojenterologii [Candidiasis of the intestine as a new problem of gastroenterology]. *Russian Journal of Gastroenterology, Hepatology, Coloproctology*, 6, 33–38 (in Russian).